Skeletal energy homeostasis: a paradigm of endocrine discovery

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Abstract

Throughout the last decade, significant developments in cellular, molecular and mouse models have revealed major endocrine functions of the skeleton. More recent studies have evolved the interplay between bone-specific hormones, the skeleton, marrow adipose tissue, muscle and the brain. This review focuses on literature from the last decade, addressing the endocrine regulation of global energy metabolism via the skeleton. In addition, we will highlight several recent studies that further our knowledge of new endocrine functions of some organs; explore remaining unanswered questions; and, finally, we will discuss future directions for this more complex era of bone biology research.

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Key Words

- marrow adipose tissue
- metabolism
- ▶ bone
- osteoblast
- ▶ pancreas

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Introduction

Bone has long been regarded as an organised collection of inert calcified structures that facilitate the motility of land animals. The skeleton's mass and composition provides vital organ protection, a niche for haematopoiesis and allows for weight-bearing motion (Guntur & Rosen 2012, Oldknow *et al.* 2015). To facilitate these classical functional roles, and to maintain bone integrity, there is a continuous homeostatic adjustment of the skeletal architecture and composition. Central to this adjustment is the highly regulated interplay of two distinct bone cell types, the osteoblast and the osteoclast, which have opposing functions (Crockett *et al.* 2011).

Osteoblasts comprise 5% of all bone cells and facilitate the formation of bone (Florencio-Silva *et al.* 2015). Mature osteoblasts synthesise and release type 1 collagen, which forms the majority (85–90%) of the organic matrix of the bone (Karsenty *et al.* 2009). Osteoblasts that become embedded in the bone matrix undergo terminal differentiation, giving rise to osteocytes – the most abundant skeletal cell type (90% of total bone cells) (Dallas & Bonewald 2010). These immobilised cells are ideally suited to perform the function of translating mechanical strain into biochemical signals in order to regulate bone composition (Sugiyama et al. 2010) (Fig. 1). The bone itself is a dynamic organ that is constantly being remodelled. This is possible due to the unique function of osteoclasts, which mediate destruction (resorption) of the bone tissue in which they reside (Holtrop & King 1977). The biphasic action of osteoblasts and osteoclasts enables bone modelling and remodelling. Bone modelling occurs throughout the lifespan, allowing the bone to adapt altered stresses and strains put on it (e.g. the tennis players serving arm), whereas bone remodelling (maintenance) occurs when the resorbed bone is completely replaced by new bone (Hadjidakis & Androulakis 2006). The regenerative process of a structure that contributes to such a large proportion of the body mass (approximately 15% in men and 10% in women) requires an abundance of proteins to be synthesised and secreted. It is therefore plausible that a high energetic cost is associated with



Figure 1

Bone anatomy and composition. Bone is organised into two distinct structures, cortical and trabecular. Cortical bone accounts for 80% of the skeletal mass and is highly organised, consisting of concentric lamellae arranged in Haversian systems. Trabecular, or 'spongy' bone, possesses ten times the surface area of cortical bone, accounting for 20% of the bone mass and enabling bone to withstand compressive and tensile forces. The bone contains osteoblasts, osteocytes and osteoclasts. Osteoblasts constitute approximately 5% of all bone cells and are the specialised 'bone-building' cells, originating from pluripotent mesenchymal stem cells (MSCs). Following matrix deposition and mineralisation, osteoblasts either remain on the surface of the bone as inactive lining cells; undergo apoptosis or become entombed by their secreted matrix and differentiate into osteocytes. Osteocytes reside within the mineralised bone matrix and are organised in functional syncytia collectively referred to as the osteocytic lacunar-canalicular system. Osteoclasts are derived from the haematopoietic lineage and are responsible for the resorption of mineralised bone and, in partnership with osteoblasts, regulate remodelling of bone tissue. The bone marrow further provides the haematopoietic niche, which supports the survival, self-renewal and differentiation of the haematopoietic stem cell (HSC). HSCs are capable of differentiation into two cell types: firstly, the common myeloid progenitor, which further differentiates to give rise to a number of blood cells including platelets, eosinophils, basophils, neutrophils, monocytes and erythrocytes; and secondly, the common lymphoid progenitor, which further differentiates to form B- and T-cells of the immune system. Within the bone marrow cavity, maintenance of the haematopoietic niche is orchestrated through vascular niches, which balance quiescence of HSC, proliferation and also regeneration following injury to the bone marrow. This regulation of HSC homeostasis involves intrinsic and extrinsic signals from the niche, including bound or secreted molecules, contractile force or even temperature. Haematological malignancies, or chemotherapy/radiation as a treatment for the disease, cause a limit to the regenerative and differentiation potentials of HCSs, causing a functional deficit (further discussed within text - see 'Disease and bone' section).

these diverse skeletal functions (Vaananen *et al.* 2000, Karsenty & Ferron 2012).

From an evolutionary perspective, bones likely represent a strongly selected survival factor that permitted enhanced movement to allow scavenging, survive injury and therefore the survival of the organism. However, it is now clear that part of the selection process for bones involves its integral role in the endocrine control of wholebody energy metabolism (Guntur & Rosen 2013). One example of the poorly understood metabolic functions of the skeleton is the presence of adipose tissue within the bone marrow – referred to as marrow adipose tissue (MAT). Accounting for approximately 10% of the total fat mass in healthy humans, the function of MAT and its association with bone-specific cells, namely osteoblasts, osteocytes and osteoclasts, remains unknown. Here, we focus on recent discoveries that explain the endocrine functions and molecular mechanisms linking bone (inclusive of MAT and muscle) and energy expenditure.

Bone as an endocrine organ

In addition to its structural role, bone is a wellrecognised target for endocrine function. This is exemplified by the orchestrated inter-organ regulation of phosphate, which involves the parathyroid glands, kidneys and intestines facilitating homeostatic maintenance of phosphate, in the mineralisation of bone extracellular matrix (Karsenty & Olson 2016). Implicit to the theory of homeostatic control is reciprocal crosstalk between the bone and these organs (Ramsay & Woods 2014). Indeed, the skeleton acts not only as an endocrine target but also as an endocrine

organ with possible roles in the hormonal modulation of systemic energy homeostasis.

Also known as BGP (bone Gla protein), osteocalcin (OCN)

Osteocalcin

is the most abundant osteoblast-specific non-collagenous protein. OCN is initially synthesised by the osteoblast as a pre-pro-molecule and is commonly used as a serum marker of bone formation (Brown et al. 1984). OCN exists in the general circulation in fully carboxylated, partially carboxylated and completely uncarboxylated forms (Plantalech et al. 1991, Cairns & Price 1994, Vergnaud et al. 1997, Schilling et al. 2005). Uncarboxylated OCN is formed when carboxylated OCN in the bone extracellular matrix is decarboxylated by the acidic pH (4.5) in osteoclastic resorption lacunae. Uncarboxylated OCN promotes β-cell proliferation, insulin secretion, peripheral insulin sensitivity and energy expenditure and impacts memory and male fertility (Lee et al. 2007, Oury et al. 2011, 2013). Recently a role for OCN in muscle function has been demonstrated. OCN levels doubled during endurance exercise in young adult wild-type (WT) mice, decreased significantly prior to or at mid-life, and OCN failed to increase during exercise in older mice. Importantly equivalent decreases in circulating OCN levels were observed in female rhesus monkeys and humans (Mera et al. 2016a). OCN administration was sufficient to reverse the age-induced decrease in exercise capacity in mice. Specifically, in 15-month-old mice, injections of OCN raised circulating OCN levels more than 4-fold and allowed these mice to run for the same time and distance as 3-month-old mice. Moreover, undercarboxylated OCN promoted uptake and subsequent catabolism of glucose and fatty acids in myofibres (Mera et al. 2016a,b). These nutrients, in turn, facilitate physical adaptation to exercise, whilst concurrently promoting the exerciseinduced release of interleukin-6 (IL-6) from skeletal muscle. IL-6 further drives the production of bioactive OCN, supporting the hypothesis of a bone-muscle feedforward axis. Thus, in addition to its postulated role in glucose and weight homeostasis (Oldknow et al. 2015), OCN further contributes to the regulation of energy metabolism, through effects on skeletal muscle. This supports the hypothesis that insulin signalling mediates

NPP1 and PHOSPHO1

In order to further increase our knowledge of the skeletons' endocrine links with energy expenditure, the role of bone mineralisation factors such as phosphoethanolamine/phosphocholine phosphatase 1 (PHOSPHO1) and ectonucleotide pyrophosphatase/ phosphodiesterase 1 (NPP1) have been addressed. NPP1, encoded by the *Enpp1* gene in mice, is highly abundant in the plasma membrane (external side) and mineral-depositing matrix vesicles of the osteoblast (Mackenzie et al. 2012). NPP1 generates inorganic pyrophosphate (PPi) through the hydrolysis of nucleotides (ATP). PPi potently inhibits hydroxyapatite crystal formation in tissues capable of mineralisation (bone and soft tissue) and acts as a precursor for inorganic phosphate (Pi) (Buckley et al. 1990, Mackenzie et al. 2012). NPP1 regulates glucose homeostasis via suppression of insulin receptor signalling in various tissues, including adipose, bone and muscle (Maddux et al. 1995, Mackenzie et al. 2012). NPP1 binds to and inhibits insulin-induced receptor conformational changes and is a potential pathogenic contributor to insulin resistance (Huesa et al. 2014). This concept is supported by the phenotype of *Enpp1* ablated mice, which display improved glucose homeostasis and resist obesity-associated dysfunction in response to high-fat diet feeding (Huesa et al. 2014). Thus, NPP1 plays multifaceted roles in normal physiology, including the regulation of calcium and phosphate homeostasis, inhibition of soft tissue mineralisation, maintenance of skeletal function and structure regulation of insulin signalling and energy homeostasis.

The bone-specific phosphatase PHOSPHO1 is a member of the large haloacid dehalogenase (HAD) superfamily of Mg2+-dependent hydrolases (Roberts et al. 2004). PHOSPHO1 is active inside the osteoblast-derived matrix vesicle, where it scavenges Pi from matrix vesicle membrane phospholipids to promote intravascular hydroxyapatite deposition. Recent studies have identified novel roles of this bone-derived factor in energy homeostasis. Mice with Phospho1 ablation exhibit a decreased body size and protection against both obesity and diabetes, regardless of carboxylation status of OCN (Oldknow et al. 2013, Chambers et al. 2015, Dayeh et al. 2016, Sayols-Baixeras et al. 2016); however, the mechanisms conferring this metabolic-protective phenotype remain to be determined.

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the link between bone remodelling, and whole-body

energy expenditure, and points towards a key role for the

osteoblast in this relationship (Huesa et al. 2014).

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PPARγ

The transcription factor PPARy is critical for differentiation of adipocytes and maintenance of the adipogenic phenotype. This is achieved via directing lineage commitment of marrow mesenchymal stem cells from an osteoblast-fate and towards that of adipocytes (Lecka-Czernik 2010). PPARy insufficiency in mice results in decreased adipose tissue and increased bone mass via inhibition of osteoclastogenesis and bone resorption (Akune et al. 2004). It remains unclear whether increased bone mass is a result of altered lineage commitment of bone marrow stem cells or an indirect effect through the modified function of adipose tissue. Alternatively, both direct and indirect mechanisms could account for the bone mass phenotype: PPARy disruption in adipose tissue (i.e. lipodystrophic disease) resulted in increased osteoblast activity and concomitant increased bone formation. The mechanisms by which PPARy regulates bone is not clear as mouse models of bone-specific PPARy conditional knockouts have not been investigated to date (Cao et al. 2015). To add further complexity, PPARy deletion in other tissues causes profound effects on bone, further complicating investigative efforts. Osteoblastselective PPARy deletion in mice (using PPAR(fl/fl):Col3.6-Cre) completely abolished adipogenesis, with the bone phenotype of increased osteoblastogenesis reflected in primary bone marrow culture and in isolated bone marrow stem cells. PPAR γ is situated at the bifurcation of lineage commitment of bone and adipocytes, suggesting that therapeutic manipulation may help to manage obesity-related conditions and orthopaedic health (Lecka-Czernik 2010). For example, rosiglitazone (an insulinsensitising thiazolidinedione) activates PPARy and effectively treats T2DM by promoting insulin sensitivity. However, rosiglitazone use comes at a cost of increased fracture risk consistent with increased adipogenesis and reduced osteoblastogenesis. With a promise for the effective management of T2DM, further work must continue to determine and thus avoid, any negative bone phenotype associated with thiazolidinedione use (Fukunaga et al. 2015).

Unexplored candidates

In light of the newly identified function of bone in energy metabolism, it is of interest to review the evidence for substrate utilisation in bone cells types. Overexpression of the glucose transporter Glut1 in osteoblasts enhances osteoblast differentiation and

http://joe.endocrinology-journals.org DOI: 10.1530/JOE-17-0147 bone formation (Wei *et al.* 2015). Assessment of glucose utilisation by the skeleton *in vivo*, using uptake of positron-emitting ¹⁸F-fluorodeoxyglucose ([¹⁸F]-FDG), revealed greater glucose uptake in bone than that in classical glucose storage and utilisation organs such as the liver, muscle and white adipose tissue (WAT) (Zoch *et al.* 2016). Furthermore, skeletal [¹⁸F]-FDG uptake was greater in young than in older mice, which may be due to the rapid bone formation in young mice. Intriguingly, insulin administration significantly increased skeletal accumulation of [¹⁸F]-FDG, whilst insulin receptor-deficient and obese mice had reduced uptake (Zoch *et al.* 2016). These findings suggest that the skeleton is a preferential and significant site of glucose uptake that is regulated by insulin and global metabolism.

Bone and adipose tissue

In times of a positive energy balance (i.e. energy intake> energy expenditure), WAT stores excess energy as triacylglycerol (TAG) and releases fatty acids (FA) and glycerol to be used for β -oxidation or gluconeogenesis during negative energy balance, respectively (Cahill 2006, Rosen & Spiegelman 2014). In addition to the role of adipose tissue in energy storage and release, adipose tissue also provides vital structural/mechanical protection for organs (e.g. the eye fat, pad, toes and heel) (Rosen & Spiegelman 2014) and offers a critical thermoprotective layer against low ambient temperatures.

Discovery of adipose-derived circulating factors such as adipsin, TNF-α, leptin and adiponectin (Badman & Flier 2007, Rosen & Spiegelman 2014) defined adipose tissue as a bona fide endocrine organ. Through the release of these 'adipokines', WAT can exert diverse systemic effects, not only on energy homeostasis but also on other aspects of physiology such as blood pressure, immune function and fertility (Michalakis et al. 2013). Thus, despite its association with metabolic diseases, WAT performs many essential physiological functions. Indeed, in the absence, and/or the redistribution of adipose tissue (lipodystrophy), patients develop insulin resistance, hyperglycemia, hypertriglyceridemia, hepatic steatosis and polycystic ovary syndrome underscoring the importance of adipose formation for normal physiological function (Cortes & Fernandez-Galilea 2015).

In contrast to white adipocytes, brown adipose tissue (BAT) is specialised for heat generation by nonshivering thermogenesis. Brown adipocytes, unlike white adipocytes, have an enrichment of mitochondria

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that express uncoupling protein-1 (UCP-1). This protein uncouples the respiratory chain, allowing protons to pass from the inner membrane space to the mitochondrial matrix without passing through ATP synthase. This causes a futile cycle: oxygen is consumed to pump protons, but the resulting chemiosmotic gradient generates no ATP and instead results in the dissipation of energy as heat (Nubel & Ricquier 2006). BAT is developmentally distinct to WAT, deriving from a distinct lineage that is shared with skeletal muscle (Rosen & Spiegelman 2014). BAT activity is relatively high in small mammals and in newborn humans, whereas BAT in adult humans is less active and is situated deep within the neck and supraclavicular region. Nevertheless, BAT in adult humans remains cold responsive, as exemplified in Scandinavian workers exposed to chronic cold (Huttunen et al. 1981). Similarly, prolonged cold exposure in rodents has shown to alter WAT cells by developing a brown fat-like morphology. These cells have been named 'beige' adipocytes, with a gene expression pattern overlapping but distinct to that of classical BAT (Wu et al. 2012, Rosen & Spiegelman 2014).

Whilst these adipose subtypes have received extensive research focus, the MAT within the marrow cavity of the skeleton has been largely ignored. Concurrent with the emergence of the field of skeletal energy homeostasis, the research into the form and function of MAT has begun to expand. Postnatally, MAT forms at distal skeletal sites, including the tailbone, hands and feet in mice and humans (Scheller & Rosen 2014). Throughout life, MAT (yellow marrow) continues to form in areas of the haematopoietic marrow (red marrow) until almost the entirety of the appendicular skeleton is converted into yellow marrow by the age of 20 years in humans (Moerman et al. 2004); however, red marrow persists in the axial skeleton, only declining with advanced age (Justesen et al. 2001). Marrow adipocytes are derived from a distinctive progenitor cell that expresses osterix, Prrx1, LepR and Gremlin1 (Chen et al. 2014). Thus, marrow adipocytes may be highly related to osteoblast precursors and play a role in bone maintenance and skeletal energy (Liu et al. 2013, Mizoguchi et al. 2014).

MAT consists of two subtypes: constitutive MAT (cMAT) and regulated MAT (rMAT). cMAT is found predominantly in the distal skeleton, giving the bone marrow a yellow appearance (Scheller *et al.* 2015). In contrast, rMAT develops much later than cMAT, in the proximal skeleton, hip, ribs and lumbar/thoracic vertebrae postnatally and consists of adipocytes interspersed with red marrow. rMAT is not necessarily formed in a normal developmental/physiological manner, instead,

rMAT seems to reflect adverse stimuli or disease states (Pichardo *et al.* 2007, Rosen & Spiegelman 2014).

Many questions remain regarding the formation and function of MAT. In animal models, MAT increases in response to the contrasting interventions of calorie restriction (CR) and high-fat diet feeding (Devlin et al. 2010, Cawthorn et al. 2014, Doucette et al. 2015). Similarly, humans with anorexia nervosa show MAT expansion (Misra & Klibanski 2013). Thus, does MAT, like WAT, play a role in regulating systemic energy homeostasis? Consistent with this possibility, is the suggestion that MAT may function as an energy reservoir for ectopic lipid, protecting skeletal osteoblasts from lipotoxicity (Gunaratnam et al. 2014), as well as secreting FA, cytokines (IL-6/1 β and TNF- α) (Caers *et al.* 2007) and adipokines (leptin and adiponectin) (Rosen et al. 2009, Cawthorn et al. 2014). Moreover, there is often a relationship between bone loss and MAT expansion, which can coincide during ageing, osteoporosis, elevated glucocorticoids and cancer treatments. This further suggests a close relationship between bone-specific cells and marrow adipocytes (Moerman et al. 2004, Georgiou et al. 2012).

The diseased state

The skeleton and associated bone-secreted factors provide a complex endocrine system that is finely orchestrated with other organs including the gut, brain, liver and kidney to ensure homeostatic balance and health. Indeed, bone-associated proteins act as a bridge to link complex pathways that bring together bone turnover, mineralisation, mineral and metabolic homeostasis. When these pathways become dysregulated, affected individuals may suffer from bone, muscle and adipose pathology (Fig. 2).

Multiple myeloma and myeloma bone disease

In the instance of multiple myeloma, affected individuals with myeloma bone disease (MBD) may experience altered bone metabolism, as a consequence of myeloma cells colonising the bone marrow (Walker *et al.* 2014, Xi *et al.* 2016). The pathophysiology of MBD is characterised by an imbalance in osteoblast and osteoclast activity. The resultant disruption of bone turnover is due to two distinct mechanisms. Firstly, engrafted myeloma cells are capable of secreting osteoclast-activating factors including, but not limited to, IL- β , TNF α and parathyroid hormone-related protein. Secondly, these engrafted cells can also

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Figure 2

Regulators of bone volume, muscle mass, subcutaneous and marrow adipose tissue. (Arrow key: Red solid – increased; green solid – decreased.) Schematic representation of the key regulators of bone volume, muscle mass, subcutaneous and marrow adipose tissue. It is interesting to highlight that both calorie restriction and glucocorticoids result in the loss of adipose tissue, muscle and bone (discussed further in the text).

interact with bone marrow microenvironment-regulating cells to further encourage secretion of osteoclastactivating factors (Roodman 2010, Terpos *et al.* 2014). By orchestrating this two-pronged 'attack', myeloma cells increase osteoclastic bone resorption. Further, several molecular mechanisms have been attributed to promoting osteoblastic reduction within MBD: Wnt-antagonists Dickkopf-1 (DKK1), runt-related transcription factor 2 (RUNX2), secreted frizzled related protein-2 (sFRP-2), transforming growth factor-beta (TGF- β), heparanase and hepatocyte growth factor (HGF) (Xi *et al.* 2016).

Such mechanisms, which compromise the normal physiological bone environment, are likely linked to energetic costs and wider metabolic consequences to the individual. Indeed, energy is expended upon the synthesis and secretion of an abundance of proteins required in the bone destruction process orchestrated by osteoclasts. Furthermore, in advanced disease states, lytic regions co-localise with elevated osteoclast activity and depressed osteoblastic activity. In accordance, the greater degree of bone acidification in osteoclastic resorption lacunae provides the conditions required to liberate the hormonally active form of OCN from the bone matrix via decarboxylation. An inverse correlation of serum decarboxylated OCN levels and the severity of MBD are reported in the literature (Bataille *et al.* 1990). Furthermore, hypercalcaemia is present at the site of bone lesions due to increased osteoclastic activity. This increased bone endocrine function represents changes to normal bone homeostasis and wider systemic and metabolic effects associated with the previously discussed roles of decarboxylated circulating OCN (i.e. increased insulin sensitivity, increased pancreatic β -cell proliferation, enhanced adipocyte secretion and reduced insulin resistance; Fig. 3).

Improved understanding of the pathogenesis of MBD has led to the identification of novel therapeutic targets. DKK1 is a key regulatory factor in the normal development of bone in adulthood, acting to inhibit osteoblastogenesis and promote differentiation of mesenchymal stem cells towards adipocytes by suppressing Wnt/beta-catenin signalling. It can be hypothesised that the associated endocrine function of the increased MAT serves to propagate myelomagenesis and tumour growth, with elevated adipocyte numbers giving secretion of free fatty acids, signalling molecules (e.g. leptin, adiponectin) and myeloma-supportive adipokines (e.g. IL-6, $TNF\alpha$). A recent study revealed that blocking of DKK1 activity (or, alternatively, the addition of DKK1 antibody) resulted in a decrease of osteolytic bone disease, with a restoration



Figure 3

Integrative model of the regulation of the new endocrine functions bone, muscle and marrow adipose tissue. (Arrow key: black solid – accepted; black dashed – speculative; red – inhibitory.) Insulin secretion from the pancreas acts upon the insulin receptor on the osteoblast, which subsequently inhibits Forkhead box protein O 1 (*FoxO1*) expression and suppresses twist basic helix-loop-helix transcription factor 2 (*Twist2*), favouring bone resorption via osteoclast activation. The adipocyte-derived hormone leptin has been shown to have two opposing roles, acting centrally to inhibit bone mass accrual and peripherally, increasing osteoblast number and activity. The acidic pH generated in the resorption lacunae decarboxylates OCN on its three glutamic acid residues (GLU13, GLU17 and GLU20), which enable it to be released from the bone matrix into the general circulation. Once circulating, OCN can regulate global energy metabolism via the stimulation of insulin secretion and β -cell proliferation in the pancreas; energy expenditure by muscle and insulin sensitivity in adipose tissue, muscle and liver. Furthermore, OCN favours hippocampal development in offspring; brain function in adults and male fertility by stimulating testosterone synthesis in Leydig cells of the testis. A bone–muscle feed-forward axis exists where systemic undercarboxylated OCN signals to myofibres favouring uptake and subsequent catabolism of glucose and fatty acids, facilitating physical adaptation to exercise and release of exercise-induced IL-6. The latter drives the production of bioactive OCN. Adiponectin release from bone marrow adipose tissue may act to indirectly increase bioactive OCN by suppressing osteoblast proliferation, potentially favouring osteoclast activity. Another possibility is that excess local OCN production is responsible, at least in part, for elevated adiponectin production from MAT; however, this remains unclear.

of increased osteoblast activity and decreased myeloma tumour burden (Qiang *et al.* 2008). The bi-directional signalling of myeloma cells and bone cells requires further investigation to determine the impacts of these interactions on bone homeostasis and tumour growth. Despite accelerated interest in the field, to date MBD (and multiple myeloma) remain incurable: it is imperative that future work is conducted to further elucidate the molecular mechanisms underlying the disruption of the bone marrow microenvironment within the framework of this complex and multifactorial disease such that novel drugs may become a feasible reality for targeted therapy for the MBD patient.

Diabetes

Globally, 642 million adults are predicted to have diabetes by 2040 (Atlas 2016). The diabetic complication of fragility fractures is of increasing importance, representing an undeniably large burden for health care systems of the world. The burden of diabetic fracture can also be considered at the individual level: fracture healing

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necessitates three energetically costly processes including inflammation, repair and remodelling (Regard *et al.* 2012). It is conceivable that the associated energetic cost of this exerts a direct effect on global energy metabolism of the affected individual, although to date, no established link of fracture burden and energy metabolism has been acknowledged.

In type 1 diabetes mellitus (T1DM), bone mineral density (BMD) - the gold standard measure for the determination of fracture risk - is decreased, a product of decreased osteoblastogenesis and increased osteoblast death (McCabe 2007, Coe et al. 2011). Conversely, BMD is increased in type 2 DM (T2DM); yet, both T1DM and T2DM patients have a significantly higher fracture risk as a complication of diabetic bone disease. compared to the general public (Janghorbani et al. 2006). This indicates a wider role of under-appreciated and undefined pathophysiological mechanisms responsible for diabetes-associated bone fragility and highlights the shortcomings of our modern day fracture risk assessment techniques. It is likely that many T2DM patients of high fracture risk go unidentified prior to fracture incidence, owing to the higher BMD associated with this class of diabetes. It remains possible that the physiological paradox of elevated BMD coinciding with increased fracture risk could well be explained by the higher prevalence of fall-associated trauma amongst diabetic patients (Gregg et al. 2002, Schwartz et al. 2002). However, it is likely that the pathophysiological mechanisms that underlie bone fragility in diabetic patients are of greater complexity than initially anticipated: even when studies include falls and associated risk factors, the association between diabetes and increased fractures remains inconclusively explained (Schwartz et al. 2002).

Suggested mechanisms of diabetic fractures include complications with hyperglycaemia, oxidative stress and glycation end-product accumulation, which compromis the properties of collagen – the most abundant of the bone proteins (Napoli *et al.* 2016). Furthermore, diabetes is associated with declining renal function, associated with lower BMD, and microvascular complications, which limit blood flow to the bone. Consequently, bones have decreased exposure to circulating bioactive hormones, including OCN, which may further contribute to skeletal fragility. These factors indicate there is a poorer quality of the bone such that there is increased fracture risk for both T1DM and T2DM, despite differences in BMD between these cohorts.

Obesity and anorexia

Our knowledge of the pathogenicity of T2DM and bone disease is further complicated by the frequent overlap of T2DM with obesity. Indeed, a long-held concept is that obesity protects against fracture risk by increasing loading of the skeleton. The increased mechanical strain in obesity is sensed and translated by osteocytes, increased BMD. However, whilst seemingly logical, this concept has recently been debunked: obesity itself is an independent risk for fracture owing to compromised quality of bones, despite non-compromised BMD (Johansson *et al.* 2014, Palermo *et al.* 2016). This confounds our attempts to understand diabetes-specific endocrine mechanisms underlying diabetic-associated skeletal fragility.

Obesity further manifests bone disease through mechanisms affecting metabolism. As both marrow adipocytes and osteoblasts likely derive from a common progenitor within the BM stroma (Chen *et al.* 2014) and that obesity promotes the differentiation of adipocytes in WAT, it is possible that obesity may also stimulate marrow adipogenesis at the cost of osteoblast differentiation. This would result in the altered quality of the obese patient's bones, even if elevated mechanical strain may be giving rise to increased BMD.

In addition, obesity is often associated with chronic inflammation. Obese individuals have an altered hormonal milieu and higher circulating levels of proinflammatory cytokines. Such cytokines may serve to modify the activity of the osteoclast receptor activator of NF-kB (RANK)/RANK-Ligand (RANKL), thereby increasing osteoclastogenesis and bone resorption. In addition, the bioavailable 25 hydroxyvitamin D3 is decreased in obese individuals, likely due to storage within the excess adipose tissue, which compromises bone mineral content (Cândido & Bressan 2014). Amongst the obese population, there is also an increase in circulating bone-anabolic hormones. This includes higher levels of pancreatic hormones (insulin, amylin and preptin) and adipose-derived factors including aromatase, leptin and resistin (Karra & Batterham 2010).

On the other end of the weight spectrum, anorexia patients also exhibit a disease-bone phenotype, with greater fracture propensity. This serious psychiatric disorder manifests in emaciation of the self-starved individual (Dede *et al.* 2014). Alongside serious weight deficit, the anorexic patient further suffers from bone structural deficits, such that the skeletal mechanical capability is impaired. These individuals

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wider endocortical diameters (Dede et al. 2014). Such microarchitectural alterations increase susceptibility to bone fragility, regardless of documented BMD values. These structural defects persist even after recovery from the disease (Dede et al. 2014). In a similar fashion to the long-suffering anorexia patient, lowcalorie intake during early stages of life (i.e. during skeletal development) results in decreased bone mass, increased fracture risk and osteoporosis in adulthood (Devlin et al. 2010). These defects are most harmful during adolescence when bone accrual is paramount for the development of peak bone mass. As previously discussed, anorexia (and caloric restriction) is associated with increased MAT (Fazeli et al. 2013, Scheller & Rosen 2014). To date, over 10 distinct animal studies have found increased MAT during states of CR or starvation, such that MAT significantly increases in the proximal femur and tibia of CR mice in comparison to the control mice (Devlin et al. 2010, Cawthorn et al. 2014). Furthermore, CR in young mice decreased serum leptin and IGF1 levels. Despite elevated bone resorption and decreased bone formation and percentage body fat, MAT was significantly increased in CR mice (Devlin et al. 2010), suggesting that increased MAT is associated with impaired skeletal maturity; however, CR in rabbits causes bone loss without MAT expansion, suggesting that the latter is not necessary for the former (Cawthorn et al. 2016). In addition to decreased circulating levels of leptin and IGF1 during CR, decreased circulating oestradiol and increased circulating FGF21, ghrelin and cortisol/corticosterone levels have also been linked to elevated BM adiposity; thus, each of these factors has been suggested as mediators of MAT expansion during CR (Thompson et al. 2004, Syed et al. 2008, Devlin et al. 2010, Shen et al. 2012, Cawthorn et al. 2014, Suchacki et al. 2016, Sulston & Cawthorn 2016). These studies highlight the possibility that MAT may be responsible for endocrine signalling such that the propensity of fracture for the anorexic sufferer is increased. One key question is whether the highly energetic cost of fracture repair, coupled with emaciated status of the anorexic individual, promotes the differentiation of skeletal stem/stromal cells towards MAT to act as an 'emergency storage' of adipocytes, and thus energy, to facilitate survival during self-starvation? If so, this likely comes at the expense of osteoblasts derived from the same skeletal progenitor, thereby further potentiating bone fragility in anorexic patients.

experience decreased cortical radius thickness and

Pancreatic disease

Given the recently acknowledged bone-pancreas loop in the regulation of glucose metabolism by insulin (Faienza et al. 2015), it is possible that pancreatic diseases such as pancreatitis or pancreatic cancer may result in altered bone homeostasis and/or endocrine function. Studies both in vitro and in vivo have revealed the osteogenic nature of insulin, promoting cell proliferation, collagen synthesis and uptake of glucose. Insulin acts on bone by binding to the insulin receptor situated on the osteoblast. Recent studies (Ferron et al. 2010, Fulzele et al. 2010) have revealed that osteoblast-specific insulin receptor knockout results in decreased osteoblast numbers and bone formation, coupled with reduced OCN activity. Patients with pancreatitis suffer from the loss of exocrine and endocrine functions via inflammatory processes that cause the destruction of the pancreas. Concomitantly, a loss of islet cells (α and β cells) results in a decrease in the release of glucoregulatory hormones (glucagon, insulin and pancreatic polypeptides). This compromised insulin release is likely to also compromise osteoblastendocrine signalling to the insulin receptor. Indeed, a study by Moran and coworkers (Moran et al. 1997) revealed that patients with pancreatic insufficiency, a product of chronic pancreatitis, exhibited osteopenia and osteoporosis, although they were unable to determine the pathological mechanisms underpinning this relationship. Furthermore, preptin, a peptide hormone cosecreted by pancreatic β cells with insulin and amylin has been shown to be anabolic to bone in vitro and in vivo (Cornish et al. 2007). During osteoporosis, preptin levels are diminished, positively correlating with BMD. It is understood that preptin is involved in the pathogenesis of osteoporosis through bone formation rather than resorption. However, further studies are required to clarify whether preptin can be a new target for treating osteoporosis by promoting bone formation (Li et al. 2013).

Liver disease

The prevalence of patients with chronic liver disease experiencing fracture is estimated at 40% (Nakchbandi 2014). As the liver coordinates many key metabolic pathways, it is unsurprising that the experience of disease within this organ results in atypical metabolism: liver disease itself is the secondary leading cause of osteoporosis. However, there is a lack of epidemiological data to support the true extent of osteoporosis amongst chronic liver

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disease sufferers (Nakchbandi 2014). The liver is central to the maintenance of health processes in the individual. For example, the liver secretes bone-health-associated factors, including IGFI and fibronectin. In health, liver-secreted fibronectin circulates prior to infiltrating the bone matrix: upon infiltration, matrix mineralisation and subsequent microarchitectural properties of bone are favourably promoted. In addition, the liver is capable of acting as a target molecule for bone-active hormones, responding with the production of various endocrine molecules including IL-6. IL-6 can act directly to activate osteoclasts or can serve to stimulate RANKL production via osteoblasts, such that osteoclasts are indirectly activated. Further, the liver is capable of metabolising bone-active molecules, including OCN, such that the period of bioavailability is reduced. Yet, in disease states, such as non-alcoholic fatty liver disease, IL-6 is upregulated as a by-product of liver injury and attempted consequential liver regeneration: this increase, in turn, promotes bone resorption by active osteoclasts. Furthermore, in chronic liver disease states, a reported 92% of patients have vitamin D deficiency: as such, calcium is liberated from the bone via osteoclastic resorption to retain homeostasis within the blood. The net result of this is the loss of bone (Nakchbandi 2014).

Perspective

The last decade has witnessed growing understanding of the skeleton's ability to act as an endocrine organ. Significant developments in cellular systems and mouse models have revealed increasingly convincing evidence in favour of the skeleton's endocrine function (Fig. 3). This adds further credence in reinforcing the importance of the skeleton for survival beyond its mechanical roles. It makes sense, from an evolutionary perspective, that the skeleton produces hormones that regulate skeletal mineralisation, cooperating with other endocrine organs to control the metabolism of phosphate and calcium.

Despite the significant advances in comprehending skeletal energy homeostasis, many questions remain unanswered. Putative investigations of other bonesecreted factors (such as NPP1 and PHOSHPO1) have revealed further candidates for links in metabolic health, including significant roles in diabetes and obesity pathology. Yet, much remains to be identified about the specific mechanisms of action and novel pathways of these new candidates with regard to skeletal and metabolic homeostasis. Continued identification of bonesecreted factors and their function will aid in answering

http://joe.endocrinology-journals.org DOI: 10.1530/JOE-17-0147 the questions of how and why bone-specific regulation of energy metabolism arose. Most recently, lipocalin (LCN2), an adipokine once thought to be exclusively secreted by adipose tissue has been shown to be an osteoblast-rich, secreted protein. LCN2 crosses the blood–brain barrier to activate the melanocortin 4 receptor, resulting in appetite suppression. Murine loss- and gain-of-function experiments demonstrated that LCN2 maintains glucose homeostasis, improve glucose tolerance and insulin sensitivity; however, more compelling human data are required to fully establish the role of LCN2 (Mosialou *et al.* 2017) (Fig. 3).

Indeed, little is also known about the role of formation and function of MAT - does MAT contribute to the global regulation of energy metabolism by the skeleton? Does MAT provide a local reservoir of energy for bone-specific cells during bone remodelling or in pathological situations? Further understanding of the mechanisms involved in this bone-metabolic axis will have many diverse implications for the management of T2DM, metabolic syndrome and other diseases of bone and adipose physiology. Such knowledge will reveal unidentified mechanisms that regulate energy homeostasis, thereby allowing development of novel pharmacological approaches for managing and treating skeletal and metabolic diseases, underscoring the need for continued research into the endocrine and metabolic functions of the skeleton.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of this review.

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References

- Akune T, Ohba S, Kamekura S, Yamaguchi M, Chung UI, Kubota N, Terauchi Y, Harada Y, Azuma Y, Nakamura K, et al. 2004 PPARgamma insufficiency enhances osteogenesis through osteoblast formation from bone marrow progenitors. Journal of Clinical Investigation 113 846–855. (doi:10.1172/JCI200419900)
- Atlas ID 2016 IDF Diabetes Atlas. International diabetes federation http://www.diabetesatlas.org/
- Badman MK & Flier JS 2007 The adipocyte as an active participant in energy balance and metabolism. *Gastroenterology* **132** 2103–2115. (doi:10.1053/j.gastro.2007.03.058)

- Bataille R, Delmas PD, Chappard D & Sany J 1990 Abnormal serum bone Gla protein levels in multiple myeloma. Crucial role of bone formation and prognostic implications. *Cancer* **66** 167–172. (doi:10.1002/1097-0142(19900701)66:1<167::AID-CNCR2820660130>3.0.CO;2-9)
- Brown JP, Delmas PD, Malaval L, Edouard C, Chapuy MC & Meunier PJ 1984 Serum bone Gla-protein: a specific marker for bone formation in postmenopausal osteoporosis. *Lancet* 1 1091–1093. (doi:10.1016/ S0140-6736(84)92506-6)
- Buckley MF, Loveland KA, McKinstry WJ, Garson OM & Goding JW 1990 Plasma cell membrane glycoprotein PC-1: cDNA cloning of the human molecule, amino acid sequence, and chromosomal location. *Journal of Biological Chemistry* 265 17506–17511.
- Caers J, Deleu S, Belaid Z, De Raeve H, Van Valckenborgh E, De Bruyne E, Defresne MP, Van Riet I, Van Camp B & Vanderkerken K 2007 Neighboring adipocytes participate in the bone marrow microenvironment of multiple myeloma cells. *Leukemia* **21** 1580–1584. (doi:10.1038/sj.leu.2404658)
- Cahill GF Jr 2006 Fuel metabolism in starvation. *Annual Review of Nutrition* **26** 1–22. (doi:10.1146/annurev.nutr.26.061505.111258)
- Cairns JR & Price PA 1994 Direct demonstration that the vitamin K-dependent bone Gla protein is incompletely gamma-carboxylated in humans. *Journal of Bone and Mineral Research* **9** 1989–1997. (doi:10.1002/jbmr.5650091220)
- Cândido FG & Bressan J 2014 Vitamin D: link between osteoporosis, obesity, and diabetes? *International Journal of Molecular Sciences* **15** 6569–6591. (doi:10.3390/ijms15046569)
- Cao J, Ou G, Yang N, Ding K, Kream BE, Hamrick MW, Isales CM & Shi XM 2015 Impact of targeted PPARgamma disruption on bone remodeling. *Molecular and Cellular Endocrinology* **410** 27–34. (doi:10.1016/j.mce.2015.01.045)
- Cawthorn WP, Scheller EL, Learman BS, Parlee SD, Simon BR, Mori H, Ning X, Bree AJ, Schell B, Broome DT, *et al.* 2014 Bone marrow adipose tissue is an endocrine organ that contributes to increased circulating adiponectin during caloric restriction. *Cell Metabolism* **20** 368–375. (doi:10.1016/j.cmet.2014.06.003)
- Cawthorn WP, Scheller EL, Parlee SD, Pham HA, Learman BS, Redshaw CM, Sulston RJ, Burr AA, Das AK, Simon BR, et al. 2016 Expansion of bone marrow adipose tissue during caloric restriction is associated with increased circulating glucocorticoids and not with hypoleptinemia. Endocrinology 157 508–521. (doi:10.1210/ en.2015-1477)
- Chambers JC, Loh M, Lehne B, Drong A, Kriebel J, Motta V, Wahl S, Elliott HR, Rota F, Scott WR, *et al.* 2015 Epigenome-wide association of DNA methylation markers in peripheral blood from Indian Asians and Europeans with incident type 2 diabetes: a nested case-control study. *Lancet Diabetes and Endocrinology* **3** 526–534. (doi:10.1016/ S2213-8587(15)00127-8)
- Chen J, Shi Y, Regan J, Karuppaiah K, Ornitz DM & Long F 2014 Osx-Cre targets multiple cell types besides osteoblast lineage in postnatal mice. *PLoS ONE* **9** e85161. (doi:10.1371/journal.pone.0085161)
- Coe LM, Irwin R, Lippner D & McCabe LR 2011 The bone marrow microenvironment contributes to type I diabetes induced osteoblast death. *Journal of Cellular Physiology* **226** 477–483. (doi:10.1002/ jcp.22357)
- Cornish J, Callon KE, Bava U, Watson M, Xu X, Lin JM, Chan VA, Grey AB, Naot D, Buchanan CM, et al. 2007 Preptin, another peptide product of the pancreatic beta-cell, is osteogenic in vitro and in vivo. American Journal of Physiology: Endocrinology and Metabolism 292 E117–E122. (doi:10.1152/ajpendo.00642.2005)
- Cortes VA & Fernandez-Galilea M 2015 Lipodystrophies: adipose tissue disorders with severe metabolic implications. *Journal of Physiology and Biochemistry* **71** 471–478. (doi:10.1007/s13105-015-0404-1)
- Crockett JC, Rogers MJ, Coxon FP, Hocking LJ & Helfrich MH 2011 Bone remodelling at a glance. *Journal of Cell Science* **124** 991–998. (doi:10.1242/jcs.063032)

- Dallas SL & Bonewald LF 2010 Dynamics of the transition from osteoblast to osteocyte. *Annals of the New York Academy of Sciences* **1192** 437–443. (doi:10.1111/j.1749-6632.2009.05246.x)
- Dayeh T, Tuomi T, Almgren P, Perfilyev A, Jansson PA, de Mello VD, Pihlajamaki J, Vaag A, Groop L, Nilsson E, *et al.* 2016 DNA methylation of loci within ABCG1 and PHOSPHO1 in blood DNA is associated with future type 2 diabetes risk. *Epigenetics* **11** 482–488. (doi:10.1080/15592294.2016.1178418)
- Dede AD, Lyritis GP & Tournis S 2014 Bone disease in anorexia nervosa. *Hormones* **13** 38–56.
- Devlin MJ, Cloutier AM, Thomas NA, Panus DA, Lotinun S, Pinz I, Baron R, Rosen CJ & Bouxsein ML 2010 Caloric restriction leads to high marrow adiposity and low bone mass in growing mice. *Journal of Bone* and Mineral Research 25 2078–2088. (doi:10.1002/jbmr.82)
- Doucette CR, Horowitz MC, Berry R, MacDougald OA, Anunciado-Koza R, Koza RA & Rosen CJ 2015 A high fat diet increases bone marrow adipose tissue (MAT) but does not alter trabecular or cortical bone mass in C57BL/6J mice. *Journal of Cellular Physiology* **230** 2032–2037. (doi:10.1002/jcp.24954)
- Faienza MF, Luce V, Ventura A, Colaianni G, Colucci S, Cavallo L, Grano M & Brunetti G 2015 Skeleton and glucose metabolism: a bonepancreas loop. *International Journal of Endocrinology* 2015 758148.
- Fazeli PK, Horowitz MC, MacDougald OA, Scheller EL, Rodeheffer MS, Rosen CJ & Klibanski A 2013 Marrow fat and bone – new perspectives. *Journal of Clinical Endocrinology and Metabolism* **98** 935–945. (doi:10.1210/jc.2012-3634)
- Ferron M, Wei J, Yoshizawa T, Del Fattore A, DePinho RA, Teti A, Ducy P & Karsenty G 2010 Insulin signaling in osteoblasts integrates bone remodeling and energy metabolism. *Cell* **142** 296–308. (doi:10.1016/j. cell.2010.06.003)
- Florencio-Silva R, Sasso GR, Sasso-Cerri E, Simoes MJ & Cerri PS 2015 Biology of bone tissue: structure, function, and factors that influence bone cells. *BioMed Research International* **2015** 421746. (doi:10.1155/2015/421746)
- Fukunaga T, Zou W, Rohatgi N, Colca JR & Teitelbaum SL 2015 An insulin-sensitizing thiazolidinedione, which minimally activates PPARγ, does not cause bone loss. *Journal of Bone and Mineral Research* **30** 481–488. (doi:10.1002/jbmr.2364)
- Fulzele K, Riddle RC, DiGirolamo DJ, Cao X, Wan C, Chen D, Faugere MC, Aja S, Hussain MA, Bruning JC, et al. 2010 Insulin receptor signaling in osteoblasts regulates postnatal bone acquisition and body composition. Cell **142** 309–319. (doi:10.1016/j. cell.2010.06.002)
- Georgiou KR, Scherer MA, Fan CM, Cool JC, King TJ, Foster BK & Xian CJ 2012 Methotrexate chemotherapy reduces osteogenesis but increases adipogenic potential in the bone marrow. *Journal of Cellular Physiology* 227 909–918. (doi:10.1002/jcp.22807)
- Gregg EW, Mangione CM, Cauley JA, Thompson TJ, Schwartz AV, Ensrud KE & Nevitt MC 2002 Diabetes and incidence of functional disability in older women. *Diabetes Care* **25** 61–67. (doi:10.2337/ diacare.25.1.61)
- Gunaratnam K, Vidal C, Gimble JM & Duque G 2014 Mechanisms of palmitate-induced lipotoxicity in human osteoblasts. *Endocrinology* 155 108–116. (doi:10.1210/en.2013-1712)
- Guntur AR & Rosen CJ 2012 Bone as an endocrine organ. *Endocrine Practices* **18** 758–762. (doi:10.4158/EP12141.RA)
- Guntur AR & Rosen CJ 2013 IGF-1 regulation of key signaling pathways in bone. *BoneKEy Reports* **2** 437. (doi:10.1038/bonekey.2013.171)
- Hadjidakis DJ & Androulakis II 2006 Bone remodeling. Annals of the New York Academy of Sciences 1092 385–396. (doi:10.1196/ annals.1365.035)
- Holtrop ME & King GJ 1977 The ultrastructure of the osteoclast and its functional implications. *Clinical Orthopaedics and Related Research* 177–196.
- Huesa C, Zhu D, Glover JD, Ferron M, Karsenty G, Milne EM, Millan JL, Ahmed SF, Farquharson C, Morton NM, *et al.* 2014 Deficiency of the

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bone mineralization inhibitor NPP1 protects mice against obesity and diabetes. *Disease Models and Mechanisms* **7** 1341–1350. (doi:10.1242/dmm.017905)

- Huttunen P, Hirvonen J & Kinnula V 1981 The occurrence of brown adipose tissue in outdoor workers. *European Journal of Applied Physiology and Occupational Physiology* **46** 339–345. (doi:10.1007/ BF00422121)
- Janghorbani M, Feskanich D, Willett WC & Hu F 2006 Prospective study of diabetes and risk of hip fracture: the Nurses' Health Study. *Diabetes Care* **29** 1573–1578. (doi:10.2337/dc06-0440)
- Johansson H, Kanis JA, Oden A, McCloskey E, Chapurlat RD, Christiansen C, Cummings SR, Diez-Perez A, Eisman JA, Fujiwara S, et al. 2014 A meta-analysis of the association of fracture risk and body mass index in women. *Journal of Bone and Mineral Research* 29 223–233. (doi:10.1002/jbmr.2017)
- Justesen J, Stenderup K, Ebbesen EN, Mosekilde L, Steiniche T & Kassem M 2001 Adipocyte tissue volume in bone marrow is increased with aging and in patients with osteoporosis. *Biogerontology* **2** 165–171. (doi:10.1023/A:1011513223894)
- Karra E & Batterham RL 2010 The role of gut hormones in the regulation of body weight and energy homeostasis. *Molecular and Cellular Endocrinology* **316** 120–128. (doi:10.1016/j.mce.2009.06.010)
- Karsenty G & Ferron M 2012 The contribution of bone to wholeorganism physiology. *Nature* **481** 314–320. (doi:10.1038/nature10763)
- Karsenty G & Olson EN 2016 Bone and muscle endocrine functions: unexpected paradigms of inter-organ communication. *Cell* 164 1248–1256. (doi:10.1016/j.cell.2016.02.043)
- Karsenty G, Kronenberg HM & Settembre C 2009 Genetic control of bone formation. *Annual Review of Cell and Developmental Biology* 25 629–648. (doi:10.1146/annurev.cellbio.042308.113308)
- Lecka-Czernik B 2010 Aleglitazar, a dual PPARalpha and PPARgamma agonist for the potential oral treatment of type 2 diabetes mellitus. *IDrugs* **13** 793–801.
- Lee NK, Sowa H, Hinoi E, Ferron M, Ahn JD, Confavreux C, Dacquin R, Mee PJ, McKee MD, Jung DY, *et al.* 2007 Endocrine regulation of energy metabolism by the skeleton. *Cell* **130** 456–469. (doi:10.1016/j. cell.2007.05.047)
- Li N, Zheng YB, Han J, Liang W, Wang JY, Zhou JR, Shen Y & Zhang J 2013 Lower circulating preptin levels in male patients with osteoporosis are correlated with bone mineral density and bone formation. *BMC Musculoskeletal Disorders* **14** 49. (doi:10.1186/1471-2474-14-49)
- Liu Y, Strecker S, Wang L, Kronenberg MS, Wang W, Rowe DW & Maye P 2013 Osterix-cre labeled progenitor cells contribute to the formation and maintenance of the bone marrow stroma. *PLoS One* **8** e71318. (doi:10.1371/journal.pone.0071318)
- Mackenzie NC, Huesa C, Rutsch F & MacRae VE 2012 New insights into NPP1 function: lessons from clinical and animal studies. *Bone* 51 961–968. (doi:10.1016/j.bone.2012.07.014)
- Maddux BA, Sbraccia P, Kumakura S, Sasson S, Youngren J, Fisher A, Spencer S, Grupe A, Henzel W, Stewart TA, et al. 1995 Membrane glycoprotein PC-1 and insulin resistance in non-insulin-dependent diabetes mellitus. *Nature* **373** 448–451. (doi:10.1038/373448a0)
- McCabe LR 2007 Understanding the pathology and mechanisms of type I diabetic bone loss. *Journal of Cellular Biochemistry* **102** 1343–1357. (doi:10.1002/jcb.21573)
- Mera P, Laue K, Ferron M, Confavreux C, Wei J, Galan-Diez M, Lacampagne A, Mitchell SJ, Mattison JA, Chen Y, et al. 2016a Osteocalcin signaling in myofibers is necessary and sufficient for optimum adaptation to exercise. Cell Metabolism 23 1078–1092. (doi:10.1016/j.cmet.2016.05.004)
- Mera P, Laue K, Wei J, Berger JM & Karsenty G 2016b Osteocalcin is necessary and sufficient to maintain muscle mass in older mice. *Molecular Metabolism* 5 1042–1047. (doi:10.1016/j. molmet.2016.07.002)

- Michalakis K, Mintziori G, Kaprara A, Tarlatzis BC & Goulis DG 2013 The complex interaction between obesity, metabolic syndrome and reproductive axis: a narrative review. *Metabolism* **62** 457–478. (doi:10.1016/j.metabol.2012.08.012)
- Mizoguchi T, Pinho S, Ahmed J, Kunisaki Y, Hanoun M, Mendelson A, Ono N, Kronenberg HM & Frenette PS 2014 Osterix marks distinct waves of primitive and definitive stromal progenitors during bone marrow development. *Developmental Cell* **29** 340–349.
- Misra M & Klibanski A 2013 Anorexia nervosa, obesity and bone metabolism. *Pediatric Endocrinology Reviews* **11** 21–33.
- Moerman EJ, Teng K, Lipschitz DA & Lecka-Czernik B 2004 Aging activates adipogenic and suppresses osteogenic programs in mesenchymal marrow stroma/stem cells: the role of PPAR-gamma2 transcription factor and TGF-beta/BMP signaling pathways. *Aging Cell* 3 379–389. (doi:10.1111/j.1474-9728.2004.00127.x)
- Moran CE, Sosa EG, Martinez SM, Geldern P, Messina D, Russo A, Boerr L & Bai JC 1997 Bone mineral density in patients with pancreatic insufficiency and steatorrhea. *American Journal of Gastroenterology* 92 867–871.
- Mosialou I, Shikhel S, Liu J-M, Maurizi A, Luo N, He Z, Huang Y, Zong H, Friedman RA, Barasch J, et al. 2017 MC4R-dependent suppression of appetite by bone-derived lipocalin 2. *Nature* (advance online publication) 543 385–390. (doi:10.1038/nature21697)
- Nakchbandi IA 2014 Osteoporosis and fractures in liver disease: relevance, pathogenesis and therapeutic implications. World Journal of Gastroenterology 20 9427–9438. (doi:10.3748/wjg.v20.i28.9427)
- Napoli N, Chandran M, Pierroz DD, Abrahamsen B, Schwartz AV & Ferrari SL 2016 Mechanisms of diabetes mellitus-induced bone fragility. *Nature Reviews Endocrinology*. **13** 208–219. (doi:10.1038/ nrendo.2016.153)
- Nubel T & Ricquier D 2006 Respiration under control of uncoupling proteins: clinical perspective. *Hormone Research* **65** 300–310. (doi:10.1159/000092847)
- Oldknow K, Morton N, Yadav M, Rajoanah S, Huesa C, Bunger L, Ball D, Ferron M, Karsenty G & MacRae V 2013 PHOSPHO1: recognition of roles beyond skeletal mineralization. In *Journal of Bone and Mineral Research.* 111 RIVER ST, Hoboken 07030-5774, NJ USA: Wiley-Blackwell.
- Oldknow KJ, MacRae VE & Farquharson C 2015 Endocrine role of bone: recent and emerging perspectives beyond osteocalcin. *Journal of Endocrinology* **225** R1–R19. (doi:10.1530/JOE-14-0584)
- Oury F, Sumara G, Sumara O, Ferron M, Chang H, Smith CE, Hermo L, Suarez S, Roth BL, Ducy P, *et al.* 2011 Endocrine regulation of male fertility by the skeleton. *Cell* **144** 796–809. (doi:10.1016/j. cell.2011.02.004)
- Oury F, Ferron M, Huizhen W, Confavreux C, Xu L, Lacombe J, Srinivas P, Chamouni A, Lugani F, Lejeune H, *et al.* 2013 Osteocalcin regulates murine and human fertility through a pancreas-bone-testis axis. *Journal of Clinical Investigation* **123** 2421–2433. (doi:10.1172/JCI65952)
- Palermo A, Tuccinardi D, Defeudis G, Watanabe M, D'Onofrio L, Lauria Pantano A, Napoli N, Pozzilli P & Manfrini S 2016 BMI and BMD: the potential interplay between obesity and bone fragility. *International Journal of Environmental Research and Public Health* **13** 544. (doi:10.3390/ijerph13060544)
- Pichardo JC, Trindade AA, Brindle JM & Bolch WE 2007 Method for estimating skeletal spongiosa volume and active marrow mass in the adult male and adult female. *Journal of Nuclear Medicine* **48** 1880–1888. (doi:10.2967/jnumed.107.044354)
- Plantalech L, Guillaumont M, Vergnaud P, Leclercq M & Delmas PD 1991 Impairment of gamma carboxylation of circulating osteocalcin (bone gla protein) in elderly women. *Journal of Bone and Mineral Research* 6 1211–1216. (doi:10.1002/jbmr.5650061111)
- Qiang YW, Chen Y, Stephens O, Brown N, Chen B, Epstein J, Barlogie B & Shaughnessy JD Jr 2008 Myeloma-derived Dickkopf-1 disrupts Wnt-regulated osteoprotegerin and RANKL production by osteoblasts:

a potential mechanism underlying osteolytic bone lesions in multiple myeloma. *Blood* **112** 196–207. (doi:10.1182/blood-2008-01-132134)

- Ramsay DS & Woods SC 2014 Clarifying the roles of homeostasis and allostasis in physiological regulation. *Psychological Reviews* **121** 225–247. (doi:10.1037/a0035942)
- Regard JB, Zhong Z, Williams BO & Yang Y 2012 Wnt signaling in bone development and disease: making stronger bone with Wnts. *Cold Spring Harbor Perspectives in Biology* 4. (doi:10.1101/cshperspect.a007997)
- Roberts SJ, Stewart AJ, Sadler PJ & Farquharson C 2004 Human PHOSPHO1 exhibits high specific phosphoethanolamine and phosphocholine phosphatase activities. *Biochemical Journal* **382** 59–65. (doi:10.1042/BJ20040511)
- Roodman GD 2010 Pathogenesis of myeloma bone disease. Journal of Cellular Biochemistry 109 283–291.
- Rosen ED & Spiegelman BM 2014 What we talk about when we talk about fat. *Cell* **156** 20–44. (doi:10.1016/j.cell.2013.12.012)
- Rosen CJ, Ackert-Bicknell C, Rodriguez JP & Pino AM 2009 Marrow fat and the bone microenvironment: developmental, functional, and pathological implications. *Critical Reviews in Eukaryotic Gene Expression* **19** 109–124. (doi:10.1615/CritRevEukarGeneExpr.v19.i2.20)
- Sayols-Baixeras S, Subirana I, Lluis-Ganella C, Civeira F, Roquer J, Do AN, Absher D, Cenarro A, Munoz D, Soriano-Tarraga C, *et al.* 2016 Identification and validation of seven new loci showing differential DNA methylation related to serum lipid profile: an epigenomewide approach. The REGICOR Study. *Human Molecular Genetics* **15** 4556–4565. (doi:10.1093/hmg/ddw285)
- Scheller EL & Rosen CJ 2014 What's the matter with MAT? Marrow adipose tissue, metabolism, and skeletal health. *Annals of the New York Academy of Sciences* **1311** 14–30. (doi:10.1111/nyas.12327)
- Scheller EL, Doucette CR, Learman BS, Cawthorn WP, Khandaker S, Schell B, Wu B, Ding SY, Bredella MA, Fazeli PK, *et al.* 2015 Region-specific variation in the properties of skeletal adipocytes reveals regulated and constitutive marrow adipose tissues. *Nature Communications* **6** 7808. (doi:10.1038/ncomms8808)
- Schilling AF, Schinke T, Munch C, Gebauer M, Niemeier A, Priemel M, Streichert T, Rueger JM & Amling M 2005 Increased bone formation in mice lacking apolipoprotein E. *Journal of Bone and Mineral Research* 20 274–282. (doi:10.1359/JBMR.041101)
- Schwartz AV, Hillier TA, Sellmeyer DE, Resnick HE, Gregg E, Ensrud KE, Schreiner PJ, Margolis KL, Cauley JA, Nevitt MC, et al. 2002 Older women with diabetes have a higher risk of falls: a prospective study. *Diabetes Care* 25 1749–1754. (doi:10.2337/diacare.25.10.1749)
- Shen W, Scherzer R, Gantz M, Chen J, Punyanitya M, Lewis CE & Grunfeld C 2012 Relationship between MRI-measured bone marrow adipose tissue and hip and spine bone mineral density in African-American and Caucasian participants: the CARDIA Study. *Journal of Clinical Endocrinology and Metabolism* **97** 1337–1346. (doi:10.1210/ jc.2011-2605)

- Suchacki KJ, Cawthorn WP & Rosen CJ 2016 Bone marrow adipose tissue: formation, function and regulation. *Current Opinion in Pharmacology* 28 50–56. (doi:10.1016/j.coph.2016.03.001)
- Sugiyama T, Price JS & Lanyon LE 2010 Functional adaptation to mechanical loading in both cortical and cancellous bone is controlled locally and is confined to the loaded bones. *Bone* **46** 314–321. (doi:10.1016/j.bone.2009.08.054)
- Sulston RJ & Cawthorn WP 2016 Bone marrow adipose tissue as an endocrine organ: close to the bone? *Hormone Molecular Biology and Clinical Investigation* **28** 21–38. (doi:10.1515/hmbci-2016-0012)
- Syed FA, Oursler MJ, Hefferanm TE, Peterson JM, Riggs BL & Khosla S 2008 Effects of estrogen therapy on bone marrow adipocytes in postmenopausal osteoporotic women. *Osteoporosis International* **19** 1323–1330. (doi:10.1007/s00198-008-0574-6)
- Terpos E, Berenson J, Raje N & Roodman GD 2014 Management of bone disease in multiple myeloma. *Expert Review of Hematology* 7 113–125. (doi:10.1586/17474086.2013.874943)
- Thompson NM, Gill DA, Davies R, Loveridge N, Houston PA, Robinson IC & Wells T 2004 Ghrelin and des-octanoyl ghrelin promote adipogenesis directly in vivo by a mechanism independent of the type 1a growth hormone secretagogue receptor. *Endocrinology* **145** 234–242. (doi:10.1210/en.2003-0899)
- Vaananen HK, Zhao H, Mulari M & Halleen JM 2000 The cell biology of osteoclast function. *Journal of Cell Science* **113** 377–381.
- Vergnaud P, Garnero P, Meunier PJ, Breart G, Kamihagi K & Delmas PD 1997 Undercarboxylated osteocalcin measured with a specific immunoassay predicts hip fracture in elderly women: the EPIDOS Study. *Journal of Clinical Endocrinology and Metabolism* 82 719–724. (doi:10.1210/jc.82.3.719)
- Walker RE, Lawson MA, Buckle CH, Snowden JA & Chantry AD 2014 Myeloma bone disease: pathogenesis, current treatments and future targets. *British Medical Bulletin* **111** 117–138. (doi:10.1093/bmb/ ldu016)
- Wei J, Shimazu J, Makinistoglu MP, Maurizi A, Kajimura D, Zong H, Takarada T, Iezaki T, Pessin JE, Hinoi E, *et al.* 2015 Glucose uptake and Runx2 synergize to orchestrate osteoblast differentiation and bone formation. *Cell* **161** 1576–1591. (doi:10.1016/j. cell.2015.05.029)
- Wu J, Boström P, Sparks LM, Ye L, Choi JH, Giang A-H, Khandekar M, Nuutila P, Schaart G, Huang K, et al. 2012 Beige adipocytes are a distinct type of thermogenic fat cell in mouse and human. Cell 150 366–376. (doi:10.1016/j.cell.2012.05.016)
- Xi H, An R, Li L, Wang G, Tao Y & Gao L 2016 Myeloma bone disease: progress in pathogenesis. *Progress in Biophysics and Molecular Biology* 122 149–155. (doi:10.1016/j.pbiomolbio.2016.08.003)
- Zoch ML, Abou DS, Clemens TL, Thorek DL & Riddle RC 2016 In vivo radiometric analysis of glucose uptake and distribution in mouse bone. *Bone Research* **4** 16004. (doi:10.1038/boneres.2016.4)

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